

Northwest
Life Science Specialties, LLC

Premier Products for Superior Life Science Research

NWLSS™ Nitrotyrosine
ELISA

Product NWK-NTR01-01
(1 plate, 96 wells)
For Research Use Only

Simple ELISA kit for quantification of nitrotyrosine in biological samples.

Note: This product contains a revised formulation and instructions for use. The Streptavidin Peroxidase reagent has been changed from a lyophilized solid to a concentrated solution.

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Introduction:

Nitrate (NO₃) and nitrite (NO₂) production are often used as markers of nitric oxide (NO) production. These analytes by themselves fail to address the eventual fate of NO or the possible adverse effects associated with its excess production and reaction with free radical species *in vivo*. Active NO metabolites can react with superoxide to form peroxynitrite (ONOO⁻) a powerful oxidant and nitrating agent. Subsequent reaction of ONOO⁻ with proteins results in nitrotyrosine (NT) formation. As a stable end product of ONOO⁻ mediated oxidation/nitration, NT can be used as a surrogate index of NO dependent damage *in vivo* and has been associated with multiple disease states.

Intended Use:

The NWLSS™ Nitrotyrosine ELISA assay is intended for the quantitative measurement of nitrosylated protein adducts in biological samples. The NWLSS™ product allows quantification of oxidative/nitrosative stress and peroxynitrite formation on proteins in biological systems.

Test Principle:

The NWLSS™ Nitrotyrosine ELISA test is a simple “sandwich” ELISA using a plate bound capture antibody (anti nitrated KLH) to nitrotyrosine and a biotinylated secondary tracer antibody. Addition of streptavidin-peroxidase followed by tetramethylbenzidine (TMB) facilitates color development directly proportional to the nitrotyrosine present in the sample. The reaction is stopped using a citric acid solution and the assay is read on a plate reader at 450 nm.

Specifications:

Format::	2 X 96 well ELISA	
Number of tests:	Triplicate =	24
	Duplicate =	40
Specificity:	Nitrosylated protein adducts	
Sensitivity:	2 nM	
Range:	2 nM—1500 nM	

Kit Contents:

40X Concentrated Wash Buffer containing Tween-20:	1 X 30 mL
10X Concentrated Dilution Buffer. Protein stabilized phosphate buffered saline containing 2-chloroacetamide preservative.	1 X 15 mL
Lyophilized Nitrotyrosine Standards Concentration is lot specific and indicated on label.	2 Vials
Lyophilized (1 mL) Biotinylated Anti-nitrotyrosine In protein stabilized buffer with 2-chloroacetamide preservative.	1 Vials
100X Streptavidin-peroxidase Reagent:	1 X 0.25 mL
TMB Substrate:	1 x 11 mL
Stop Solution containing oxalic acid:	1 X 22 mL
96 well microplates (12 X 8 well strips) pre-coated with anti-nitrotyrosine antibody:	1 each

Required Materials Not Provided:

Adjustable micropipettes with disposable tips (50-1000 μ L). Multi-channel pipettes are useful and help to reduce intra-sample variability.

Serological pipettes.

Deionized water.

Polypropylene tubes

Automatic plate washer or other aspiration devices are optional.

Required Instrumentation:

Plate reader with 450 nm capability.

Warnings, Limitations, Precautions:

Do not add sodium azide as preservative to any component since its presence will inactivate the peroxidase conjugate.

Components containing 2-chloroacetamide and citric acid may be hazardous if in direct contact with skin, eyes, etc. Contact should be minimized through the use of gloves and standard good laboratory practices. If contact occurs, rinse the site immediately with water.

Storage Instructions:

Upon receipt, store this product at 2-8 °C...**DO NOT FREEZE**. Lyophilized components are stable for 1 month after reconstitution if stored at 2-8°C.

All reagents should be brought to room temperature (18-25°C) prior to use and stored at 2-8°C immediately after use.

100X SAP Conjugate is stable up to expiration date when stored at 2-8°C . Working SAP Conjugate must be used the same day it is diluted. It is not stable when stored in diluted form.

Prolonged exposure of kit components to light should be avoided.

Coated microwell strips may be used until the product expiration date as long as they are returned to pouch and stored dry at 2-8°C.

Assay Preparation

1. Determine the number of wells required to assay standards, samples and controls for the appropriate replicate. It is recommended that testing be performed in duplicate or triplicate if possible.
2. Create an assay template showing positioning of standards, controls and samples.
3. Bring all samples and reagents to room temperature before use.
4. To avoid condensation, do not open foil pouches containing the microtiter strips until after they have reached room temperature. Next remove the required number of strips and place in the frame supplied.

Note: The bottom of some wells may be slightly white due to the preservation treatment. This does not influence assay performance.

Return unused wells to the storage bag with desiccant, seal and store at 2-8°C.

Reagent Preparation:**1. 40X Wash Buffer:**

For each 96 well plate to be assayed, dilute 15 mL of 40X Wash Buffer with 585 mL de-ionized water. If assaying less than a full plate make up only the required amount. Label as **Working Wash Buffer**.

2. 10X Dilution Buffer:

For each 96 well plate to be assayed dilute 7.5 mL of 10X Dilution Buffer with 67.5 mL de-ionized water. If assaying less than a full plate make up only the required amount. Label as **Working Dilution Buffer**.

Note: Lyophilized components are under vacuum. Allow pressure to equalize slowly before opening fully.

3. Lyophilized Standards

Reconstitute each Standard vial with **Working Dilution Buffer** according to the directions on the vial label to create a 1500 nM stock solution which will also function as the high standard. Label as **Tube 1 : 1500 nM NTR**.

4: Lyophilized, Biotinylated Anti-nitrotyrosine Tracer:

Reconstitute 1 vial with 1 mL *deionized water* per 96 well plate assayed. Next dilute the 1 mL reconstituted Tracer with 11 mL *Working Dilution Buffer* and/or for each 8 well strip to be assayed dilute 83 μ L reconstituted Tracer with 917 μ L **Working Dilution Buffer**.

5. 100X Streptavidin-Peroxidase (SAP) Reagent:

Spin down this vial prior to use. For 96 wells mix 100 μ L of 100X SAP Conjugate with 9.9 mL *Working Dilution Buffer*. If less than a full plate will be assayed prepare the required volume by mixing 1 part *100X SAP Reagent* with 99 parts *Working Dilution Buffer*. Label as **Diluted SAP Conjugate** and use the same day.

6. TMB Substrate: and Stop Solution are supplied ready to use.

Standard Curve Preparation::

1. Label microtubes 2-8. Add 350 μ L **Working Dilution Buffer** to each.

2. Transfer 175 μ L of **1500 nM NTR** standard to tube 2 and mix well to create 500 nM standard. Continue 1/3 serial dilutions across tubes 3-7 creating standards of 166.7 - 2.1 nM. Leave tube 8 as a dilution buffer only zero control.

Std Tube # :	<u>1</u>	<u>2</u>	<u>3</u>	<u>4</u>	<u>5</u>	<u>6</u>	<u>7</u>	<u>8</u>
Conc. (nM):	1500	500	166.7	55.6	18.5	6.2	2.1	zero control

Standards cannot be stored for future use.

Sample Handling/Preparation

1. Before performing the assay, all samples should be brought to room temperature.
2. Samples should be diluted if necessary with Working Dilution Buffer and mixed gently taking care to avoid foaming.

Note that 100 μ L sample is required per replicate well.

Plasma or Serum samples:

1. Blood samples should be stored on ice prior to separation.
2. For plasma, EDTA anticoagulant is recommended.
3. Samples frozen long term at -80°C are suitable for assay however sample storage at -20°C may adversely affect recovery of the NT analyte.
4. Avoid multiple freeze thaw cycles. In the case of frozen samples, use thawed sample within 24 hours.
5. All samples should be handled according to standard guidelines for preventing transmission of blood borne pathogens.
6. It is recommended that plasma samples be diluted 10X prior to assay.

Tissue samples:

Tissue samples can be homogenized in PBS, pH 7.3. Homogenates should be kept as concentrated as possible then tested at 1/2, 1/5, 1/10 etc. to determine the best dilution for the specific tissue type.

Assay Protocol:

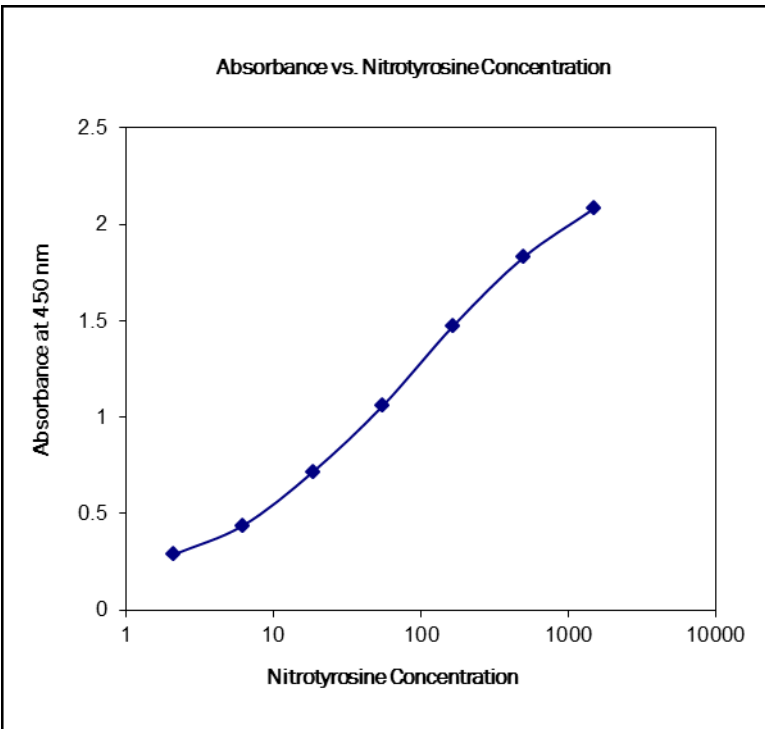
Allow approximately 4.5 hours for procedure.

1. Add **100 µL standard, sample or control** to each well according to the assay template created earlier.
2. Cover the wells using adhesive tape or plastic wrap and incubate for 1 hour at room temperature (18-25°C).
3. Carefully remove the plate cover. Empty the plate by inverting and shaking contents over sink. Keep inverted and tap dry on a thick layer of tissues or paper towels. Wells may also be aspirated using a multi-channel pipette or plate washer. In either case, wash the plate 4 times with 200 µL Working Wash Buffer waiting 20 seconds before aspirating.
5. Add **100 µL of Diluted Tracer** to each well keeping the same sequence as used in step 1. Avoid touching the side or top of the wells.
6. Cover the wells and incubate for 1 hour at room temperature.
7. Repeat washing procedure as above.
8. Add **100 µL of Diluted SAP Conjugate** to each well keeping the same sequence as used earlier.
9. Cover the wells and incubate for 1 hour at room temperature.
10. Repeat washing procedure as above.
11. Add **100 µL of TMB Substrate Solution** to each well, keeping the same sequence as used in earlier steps.
12. Cover the wells and incubate in the dark for 20-30 minutes at room temperature. Color development should be monitored to avoid over-development of high standards. Reaction can be stopped sooner if necessary.
13. Stop the reaction by adding **100 µL Stop Solution** taking care to use the same sequence and timing as previous steps.
14. Measure the absorbance at 450 nm.

The mean absorbance for the zero buffer only control should be < 0.3

Data Analysis:

1. Plot the mean absorbance at 450 nm for each standard replicate versus the nitrotyrosine concentrations setting x-axis to log scale. We recommend using a 4-parameter curve fit. This can typically be done using the software provided with most modern plate readers. An example curve is shown below.
2. Unknown nitrotyrosine concentrations are determined by comparing their absorbance measurements at 450 with those of the standard curve.



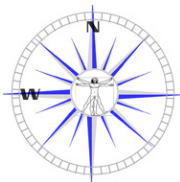
Procedure Checklist

- ___ Create an assay template
- ___ Equilibrate reagents to room temperature.
- ___ Set-up required number of strips in frame supplied.
- ___ Prepare reagents
 - Reconstitute lyophilized components
 - Dilute Reagent Vials 1, 2, 4 and 5 as required.
- ___ Perform a 1/3 serial dilution of the 1500 nM standard
- ___ Prepare samples, making any necessary dilutions.
- ___ Add 100 μ L standard, sample or control to each replicate well according to assay template.
- ___ Cover and Incubate 1 Hour at room temperature.
- ___ Wash wells 4 times with 200 μ L Working Wash Buffer
- ___ Add 100 μ L Diluted Tracer to each well.
- ___ Cover and Incubate 1 Hour at room temperature.
- ___ Wash wells 4 times with 200 μ L Working Wash Buffer
- ___ Add 100 μ L of diluted Streptavidin-Peroxidase Conjugate to each well.
- ___ Cover and Incubate 1 Hour at room temperature.
- ___ Wash wells 4 times with 200 μ L Working Wash Buffer
- ___ Add 100 μ L TMB Substrate Solution to each well.
- ___ Cover and incubate 20-30 minutes at room temperature in the dark.
- ___ Stop the reaction by adding 100 μ L Stop Solution.
- ___ Measure absorbance at 450 nm
- ___ Analyze data using standard curve plot.

Statement of Limited Warranty:

Northwest Life Science Specialties, LLC (NWLSS) makes no guarantee of any kind, expressed or implied, that extends beyond the description of the material in this kit, except that they will meet our specifications at the time of delivery. Customer's remedy and NWLSS' sole liability is limited to, at NWLSS' option, refund of the purchase price, or the replacement of material not meeting our specification. By acceptance of our product, customer assumes all liability and will indemnify and hold NWLSS harmless for the consequence of this product's use or misuse by the customer, its employees, or others. Refund or replacement is conditioned of customer notifying NWLSS within twenty-one (21) days of the receipt of product. Failure to give notice within 21 days shall constitute a waiver by the customer of all claims hereunder with respect to said product.

Notes:



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